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REGULAR ARTICLE

Fragmentation modulates the strong impact of habitat quality and plant cover on fertility and microbial activity of semiarid gypsum soils

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13 Abstract

Background and aims Plant-soil interactions are a cru-14cial component of ecosystem functioning. However, 15most global change studies focus on plant communities, 16with information on soil properties and performance 17being scarce. Our goal was to assess the individual and 18joint effect of habitat heterogeneity and three global 1920change drivers (fragmentation, loss of habitat quality and climate change) on nutrient availability and soil 21microbial activity in Mediterranean gypsum soils. 22

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T. E. Gimeno · A. Escudero · F. Valladares Departamento de Biología y Geología, ESCET, Universidad Rey Juan Carlos, Tulipán s/n, 28933 Móstoles, Spain Methods We collected soil samples from an experimen-23tal field site from large/small fragments, with high/low 24habitat quality, subjected to two levels of water avail-25ability (dry/mesic) and from two microhabitats (under 26the canopy of shrubs and in the open). We analyzed 27nutrient concentrations (C, N and P) and enzymatic 28activities (ß-glucosidase, urease and acid phosphatase). 29Results C, N, P content, ß-glucosidase, urease and 30 acid phosphatase activities were higher under the can-31opy than in the open and in high- than in poor- habitat 32quality sites. These differences were exacerbated in 33 small fragments. 34

Conclusions The strong interdependence between plant 35and soil was modulated by fragmentation in the Medi-36 terranean gypsum soils studied. Drought did not exert a 37 direct negative effect on soil properties, although the 38 effect might arise under more intense drought or under 39drought taking place at times of the year different from 40 those explored here. Results highlight the importance of 41 considering several drivers simultaneously to forecast 42realistic ecosystem responses to global change. 43

Keywords Enzymatic activities · Global change ·	44
Habitat quality · Fragmentation · Gypsum soil ·	45
Mediterranean ecosystem	46

Introduction

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Soil nutrient availability is one of the most important 48 factors influencing plant growth and ecosystem 49

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functioning (Lambers et al. 1998). The distribution of 50nutrients in the soil is highly heterogeneous, which in 5152turn affects the structure and composition of plant communities (Kruger 1979; Henkin et al. 1998). 53Moreover, several studies have shown that soil hetero-5455geneity can modulate the impact of global change drivers on plant communities (Maestre and Reynolds 562006; Wacker et al. 2008). Soil nutrient heterogeneity 57is also associated with microbial activity (Gallardo and 58Schlesinger 1994), which in turn are responsible for 59essential processes in the ecosystem such as energy 60 transformation, mineralization of plant litter and nutri-61 ent cycling (Panikov 1999). In agreement with this, 62 recent studies have evidenced the importance of con-63 sidering the below-aboveground interactions of the 64 biota to properly understand ecosystem functioning 65 (van der Putten et al. 2009; Kardol and Wardle 2010; 66 Garcia-Palacios et al. 2011). 67

Climatic conditions such as soil and air temperature 68 and water availability affect enzyme activity through 69 70increased microbial growth and substrate availability (Nov-Meir 1973; Parkinson and Coleman 1991). Sev-71eral studies have shown the importance of water avail-7273 ability for both microbial activity (Kramer and Green 742000; Li and Sarah 2003; Sardans and Penuelas 2005) and soil nutrient availability (Jensen et al. 2003; Sardans 75and Penuelas 2004). Consequently, changes in temper-7677ature or precipitation promoted by climate change are likely to alter nutrient cycles (Sardans and Penuelas 782007) and nutrient availability for plants (Michelsen et 79al. 1999). This becomes especially important in Medi-80 terranean ecosystems, where global circulations models 81 forecast reductions in precipitation and an increase in 82 maximum temperatures together with heavier storms 83 (Christensen 2007). Higher temperatures will further 84 85 decrease soil water availability and exacerbate the effects of drought in these environments (Larcher 86 2000), while heavy storms increase nutrient loss by 87 88 lixiviation (Reynolds et al. 2004), and increased runoff decreases water infiltration (Wainwright 1996). 89

90 Besides climate change, other global change drivers such as land use changes and habitat fragmentation can 91 have dramatic effects on microbial and enzymatic activ-92ity and nutrient availability (Matias et al. 2010). Medi-9394terranean ecosystems have been profoundly transformed over centuries due to human activities such 95as farming or agriculture (Valladares et al. 2008). These 96 97 have caused fragmentation and reductions in habitat quality, important threats for biodiversity and natural 98

resources conservation (Lavorel et al. 1998; Foley et 99al. 2005). Fragmentation decreases plant population size 100and increases isolation, which can lead to lower genetic 101 variability and lower individual fitness and plant surviv-102al (Lienert 2004; Aguilar et al. 2006). As a consequence, 103soils in fragmented landscapes may have reduced or-104 ganic inputs and thus reduced nutrient availability and 105cycling (Garcia et al. 2002). Reduced habitat quality has 106 Q2 often been considered a result of habitat fragmentation 107(Harrison and Bruna 1999; Schleuning et al. 2008). 108However, in agricultural landscapes, changes in habitat 109 quality may occur independently from fragmentation, 110through factors such as runoff and fertilizer drift into 111 adjacent areas, intense ploughing, trampling or soil ero-112sion (Boutin and Jobin 1998; Matesanz et al. 2009). 113Reduced habitat quality has also been associated to 114 decreased plant cover and biological soil crust, which 115is translated into a meagre input of dead organic matter 116and a consequent decrease of microbial activities (Zak et 117al. 1994). However, the direct effect of habitat fragmen-118tation and reduced habitat quality on soils attributes and 119performance remains largely unknown. 120

Interactions among global change drivers frequent-121ly generate non-additive effects, which in turn either 122attenuate or exacerbate ecosystem responses to indi-123vidual drivers (Zavaleta et al. 2003; Matesanz et al. 1242009). Several studies have addressed the interacting 125effects of global change drivers on ecosystems, but 126most of them have focussed on their influence on plant 127communities (Sala et al. 2000; Maestre and Reynolds 1282006; Matesanz et al. 2009), while information on 129microbial communities and soil nutrient availability 130is particularly scarce (Cookson et al. 2007; Casals et 131al. 2009; Matias et al. 2010). 132

Our main goal was to assess the individual and joint 133effects on nutrient availability and soil microbial activity 134of three global change drivers that are especially impor-135tant for Mediterranean ecosystems: habitat fragmenta-136tion, loss of habitat quality and water availability. 137Moreover, we assessed the influence of microhabitat 138heterogeneity (i.e. open vs. the understory of woody 139plants) and its interaction with these global change 140drivers on the same microbial and soil properties. We 141conducted a field experiment in a Mediterranean gyp-142sum steppe with plots following a factorial design for 143the three drivers. Our working hypotheses were: (1) 144 Habitat fragmentation, loss of habitat quality and reduc-145tions in rainfall decrease plant survival and productivity 146 which are strongly related to soil attributes and 147

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performance (Garcia et al. 2002; Zak et al. 2003). This 148in turn, will exert a negative effect on nutrient availabil-149ity and microbial activity of Mediterranean gypsum 150soils; (2) synergistic interactions among drivers will 151amplify the negative impact of loss of habitat quality 152on soil nutrient availability (Matias et al. 2010); and (3) 153nutrient availability and microbial activity will increase 154under the canopy of shrubs in comparison with open 155interspaces and this microhabitat heterogeneity will 156modulate the influence of other global change drivers 157as suggested on other plant communities (Reich et al. 1582001; Maestre and Reynolds 2006). 159

160 Materials and methods

161 Study site

162The study was carried out near Belinchón in central Spain (745 m above sea level; 40° 03' N, 3° 03' O). 163The landscape is composed by gypsum soil hills (av-164erage slope was $11.7\pm0.3^{\circ}$) with remnants of natural 165vegetation interspersed in a matrix of dry-farm crops. 166 Natural vegetation is dominated by creeping and 167 168cushion-like chamaephytes such as Centaurea hyssopifolia Vahl. (Compositae), Helianthemum squama-169tum (L.) Dum. Cours (Cistaceae), Lepidium 170subulatum L. (Cruciferae), Thymus lacaitae Pau 171(Labiatae) and Teucrium pumilum L. (Labiatae). Plant 172cover is usually low (<30%), and bare soil areas are 173often covered by a conspicuous biological soil crust, 174dominated by specialised lichens (Martinez et al. 1752006). The area has a Mediterranean semiarid climate, 176with a mean annual precipitation of 433 mm, a pro-177nounced summer drought, and a mean annual temper-178ature of 13.8°C. The study was conducted over 1792 years: 2005, which was the second driest year of 180the 56-year series (298 mm annual precipitation), and 1811822006, also a drier-than-average year, with annual precipitation of 371 mm (see detailed precipitation data of 183184 the study site in Online resource 1)

185 Experimental design and soil sampling

To test the effects of three global change drivers and
their interactions on soil features and performance and
to explore the effect of microhabitat, we conducted an
experiment with four controlled factors: fragmentation,
habitat quality, water availability and microhabitat. For

each factor two levels were selected: large (L) and small 191(S) fragments, high (H) and poor (P) habitat quality, 192mesic (M, watered plants) and dry (D, non watered 193 plants). Two microhabitats were considered for each 194combination of factors, under the understory of C. hys-195sopifolia (U. Understory) and open areas near the target 196 plants (O, Open). We selected this plant species because 197 it is the largest and most abundant chamaephyte in the 198 local community. 199

To select the two levels of fragmentation we identified 200three small (area <1.5 ha) and three large (area >11 ha) 201 fragments of natural vegetation (six fragments total) 202which were further characterized by measuring several 203 vegetation attributes such as percentage of soil covered 204 by plants, lichens and mosses, annual plants, perennial 205plants, litter and bare soil (see Online resource 2). Within 206each fragment, we randomly selected two plots of ca. 207 15×15 m of contrasting high- and poor-habitat quality 208 (12 plots in total) according to plant cover as an integra-209tive indicator of habitat suitability (see Matesanz et al. 2102009 for a detailed characterisation of each habitat qual-211ity level). Each plot was further divided into two contig-212uous halves that were randomly assigned to one watering 213treatment. The irrigation experiment was conducted in 214the spring (May and June) of 2005 and 2006, simulating 215two different scenarios of water availability: non-216watered plants (dry treatment) and watered plants (mesic 217treatment). Water was added to reach the median of the 218long-term series (1948-2004) in each month (Fig. 1). 219Plants were randomly selected within the mesic plot. 220Irrigation was then applied at the plant-level and con-221sisted of adding 11 of dechlorinated tap water per plant 222and application time. A 50×50 cm (0.25 m²) rigid frame 223was placed around each watered plant so that the entire 224surface was watered and all the plants received the same 225amount of water, independently of their size. Each water 226 application was equivalent to 4-mm rainfall events. Irri-227gation was performed at 5-6 days intervals. The non-228watered (dry treatment) plants received ambient precip-229 itation (equivalent to future drier scenarios due to the 230very dry spring conditions of the study years) and the 231irrigated plants received ambient precipitation plus the 232added water (equivalent to a typical year). 233

In July 2006, we randomly selected five plants per irrigation treatment and we collected soil samples from each microhabitat. The total number of soil samples was 240 (10 plants per plot x 12 plots x 2 microhabitats). We collected four sub-samples within the perimeter where the irrigation treatment was carried 239

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out with a $6 \times 6 \times 10$ cm metal soil core for each subsample, which were thoroughly mixed afterwards.

sample, which were thoroughly mixed afterwards.Once in the laboratory, soil samples were sieved

243 (2 mm grain) and air dried.

240 (2 min grain) and an dife

244 Biochemical and microbiological analysis

245Total nitrogen (N) and total phosphorous (P) contents were determined by the Kjeldahl method (Radojevic 246and Bashkin 1999). Each soil sample was digested in 24796% sulphuric acid for 3 h at 415°C and nutrient 248contents were determined through colorimetry by an 249250automatic wet chemistry analyzer (Skalar 4000 SAN System, Segmented Flow Analyzer; Skalar, Breda, 251The Netherlands). Total organic carbon (C) was deter-252mined by Walkley and Black method (1934) modified 253by Yeomans and Bremmer (1989) by oxidation with 254potassium dichromate in acid medium and evaluating 255256the excess of dichromate with 0.5 N ferrous ammonium sulphate. 257

258β-glucosidase and acid phosphatase activities were 259estimated using Tabatabai method (1982), which determined colorimetrically the amount of p-nitrophenol 260261produced from p-nitrophenyl- β-D-glucopyranoside, 262and p-nitrophenyl-phosphate, respectively, after 1 h of incubation at 37°C. The activities are expressed as 263grams of p-nitrophenol per gram of soil and hour 264(Moreno et al. 2003). Urease activity was determined 265colorimetrically by Nannipieri method (1980) measur-266 ing total ammonium produced from a buffered urea 267solution. 268

269 Statistical analysis

The effects of the different fixed factors (fragmenta-270271tion, habitat quality, water availability and microhabitat) on the dependent variables (total organic C, total 272273N, total P, β-glucosidase, urease and acid phosphatase 274activity) were analyzed using a four-way nested ANOVA model. The model included fragmentation 275276(F, 1 df), habitat quality (Q, 1 df), water availability (W, 1 df) and microhabitat (MH, 1 df) as main fixed 277factors. Each sampling point was considered as a 278279random factor nested within fragmentation level (sampling point (F), 4 df). We tested main effects of these 280fixed factors and also included all possible interactions 281282between them. When significant interactions between two factors were found, we performed a one-way 283ANOVA to test for significant effects of one factor 284

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within each level of the second factor. Normality and
homogeneity of variance in the dependent variables285was tested prior to analyses by means of the
Kolmogorov-Smirnov and the Levene's test. All sta-
tistical analyses were performed using Statistica 6.0289(StatSoft Inc., Tulsa, OK, USA).290

Results

Soil nutrients

Total organic carbon, total N and total P were signif-293icantly higher in high quality habitats and under the294understory of C. hyssopifolia (Fig. 2, Table 1). Loss of295habitat quality had the strongest impact. Fragmenta-296tion and water availability had no significant direct297effects on total organic C, N and P.298

We found significant interactions between factors 299affecting all nutrients. The interaction between habitat 300 quality and fragmentation had a significant effect on 301 organic C (Table 1, Fig. 4a) and total N (Table 1, 302 Fig. 4b). Organic C was lower in small than in large 303fragments in poor habitat quality plots (F=8.319 p=304 0.005), but not in high habitat quality plots (F=0.299, 305 p=0.586). Total N did not differ significantly between 306 large and small fragments neither in high habitat qual-307 ity (F=3.295, p=0.072), nor in poor habitat quality 308plots (F=3.451, p=0.066). The interaction between 309 habitat quality and microhabitat had a significant ef-310 fect on total N (Table 1, Fig. 4c): total N did not differ 311between open and understory in high habitat quality 312plots (F=3.237, p=0.075), but it was significantly 313lower in poor habitat quality plots (F=21.875, p<3140.001). Finally, total P was affected by a significant 315interaction between habitat quality and water avail-316 ability (Table 1, Fig. 4d), but we did not find signifi-317cant differences between watering treatments within 318levels of habitat quality (F=0.340, p=0.560; F=319 4.248, p=0.061 for high- and low-habitat quality, 320 respectively). 321

Soil enzymatic activity

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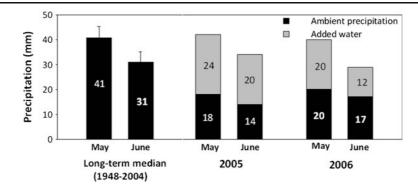


Fig. 1 Irrigation experiment. May and June precipitation medians (1948–2004 series) were used as a threshold for the irrigation treatment. Plants in the dry treatment received ambient

activity of enzymes decreased in poor habitat quality
plots, and it was lower in the open than under the
understory (Table 1, Fig. 3). We found no significant
main effects of fragmentation and water availability on
β- glucosidase, phosphatase and urease activity.

precipitation, and plants in the mesic treatment received ambient precipitation plus added water (through 4 mm events and up to the median for the corresponding month)

Phosphatase activity was significantly affected by the 333 interaction between habitat quality and water availability (Table 1, Fig. 4e). Yet, we did not find significant 335 differences between watering treatments within levels of 336 habitat quality (F=1.513, p=0.221; F=4.141, p=0.064, 337

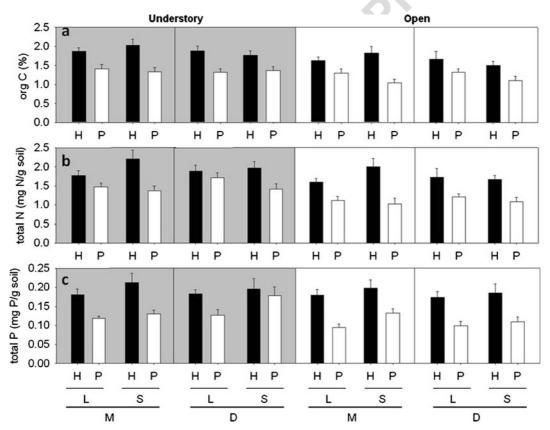


Fig. 2 Soil nutrient content across treatments. **a** Total organic C; **b** total N; **c** total P. Each half of a panel corresponds to data from understory (*left*) and open (*right*) microhabitats. Values are mean \pm SE in each treatment. Different colours indicate significant differences between microhabitats (background color) and

between high and poor habitat quality (bar colours). Abbreviations are: H, high-habitat quality; P, poor-habitat quality; L, large fragment; S small fragment; M, mesic treatment (watered plants); D, dry treatment (non-watered treatments)

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t1.1 **Table 1** ANOVA results (F and *p*-values) for the soil nutrient contents and soil enzymatic activity. N=240 soil samples. NS: not significant. See results section for direction of the effects. Significant effects (p < 0.05 are indicated in bold)

	Total organic C		Total N		Total P		β- glucosidase		Phosphatase		Urease	
	F	р	F	р	F	р	F	р	F	р	F	р
Fragmentation (F)	0.561	NS	0.002	NS	0.324	NS	0.642	NS	3.214	NS	2.850	NS
Habitat quality (Q)	56.560	0.000	70.872	0.000	92.861	0.000	20.160	0.000	4.575	0.034	0.045	NS
Water availability (W)	0.045	NS	0.259	NS	1.257	NS	1.005	NS	0.668	NS	2.482	NS
Microhabitat (MH)	19.750	0.000	23.658	0.000	7.870	0.005	65.711	0.000	26.369	0.000	20.809	0.000
$\mathbf{Q} \times \mathbf{F}$	6.056	0.015	9.393	0.002	0.347	NS	3.505	NS	2.554	NS	4.214	0.041
$\mathbf{F} \times \mathbf{W}$	2.121	NS	1.865	NS	0.576	NS	3.260	NS	1.642	NS	1.085	NS
$\mathbf{Q} imes \mathbf{W}$	3.315	NS	2.902	NS	4.735	0.031	0.251	NS	6.214	0.013	0.169	NS
$F \times MH$	0.109	NS	0.000	NS	0.290	NS	0.011	NS	0.361	NS	0.286	NS
$\mathbf{Q} \times \mathbf{M} \mathbf{H}$	0.008	NS	4.024	0.046	2.216	NS	0.597	NS	0.122	NS	0.153	NS
$W \times \mathbf{M} H$	0.949	NS	0.098	NS	0.284	NS	0.610	NS	0.462	NS	2.436	NS
$F\times Q\times W$	0.214	NS	0.596	NS	3.195	NS	0.664	NS	0.594	NS	0.431	NS
$F \times Q \times MH$	0.132	NS	0.088	NS	0.000	NS	0.342	NS	2.435	NS	1.796	NS
$F\times W\times MH$	0.048	NS	0.175	NS	0.032	NS	0.048	NS	0.102	NS	0.003	NS
$Q\times W\times MH$	0.075	NS	0.039	NS	0.238	NS	0.017	NS	0.176	NS	0.158	NS
$F \times Q \times W \times MH$	1.467	NS	0.071	NS	0.354	NS	0.005	NS	0.099	NS	0.262	NS
Sampling point(F)	4.291	0.002	10.440	0.000	85.840	NS	13.757	0.000	1.622	NS	14.997	0.000

338 for high- and low-habitat quality, respectively). Urease activity was also affected by the interaction between 339 habitat quality and fragmentation (Table 1, Fig. 4f): 340 the difference between large and small fragments 341was greatest in poor habitat quality plots. Urease 342activity was greater in small fragments, both under 343 poor (F=7.227, p=0.008) and under high habitat 344quality (F=28.861, p<0.001). 345

346 Discussion

347 Effects of habitat quality and habitat heterogeneity

348 As expected, total organic carbon, N and P, βglucosidase and acid phosphatase activities were sig-349350nificantly reduced in open interspaces and in low quality habitat sites. The relative influence of fragmen-351tation, water availability and habitat quality was dif-352ferent with a maximum impact associated with habitat 353degradation. The reduction of aboveground plant pro-354ductivity in poor quality habitats underlies reduced 355 356 organic C inputs, the main energy source for heterotrophic microbial communities (Zak et al. 2003; Allen 357 and Schlesinger 2004). This result agrees with 358

previous studies showing that microbial community 359 composition and function depend directly on plant 360 cover and soil organic matter content (Zak et al. 361 1994; Garcia et al. 2002). Limited nutrient input also 362 explains the decrease in N and P content and conse-363 quent decrease in β-glucosidase and phosphatase ac-364 tivities. These results suggest that plant abundance 365 significantly affects soil microorganisms and the eco-366 system processes they mediate, like nutrient cycling 367 (Schlesinger and Pilmanis 1998; Stephan et al. 2000; 368 Tilman et al. 2001; Zak et al. 2003). Given that soil 369 nutrient deficiencies limit plant growth (Henkin et al. 370 1998; Fenner 2001; Sardans and Penuelas 2004), we 371can expect reduced enzymatic activity to indirectly 372affect plant growth, highlighting the strong interde-373 pendence between plant and microbe soil communi-374ties, which involves positive feedbacks. 375

Microhabitat heterogeneity played an important 376 role for soil properties, affecting both nutrient content 377 and soil enzymatic activities. Higher enzymatic activ-378 ity underneath the canopy of C. hyssopifolia and in 379 high-quality sites may be due to the larger microbial 380 and root biomass densities beneath the plants, which 381entails a faster nutrient intake and stimulates the syn-382 thesis and excretion of enzymes (Garcia et al. 2002; 383

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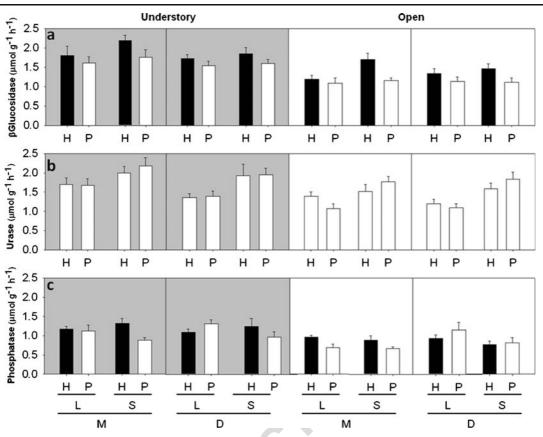


Fig. 3 Soil enzyme activity across treatments. a β -Glucosidase; b Urease; c Phosphatase. Values are mean \pm SE in each treatment. Each half of a panel corresponds to data from understory and open microhabitats. Different colours indicate significant differences between microhabitats (background color) and

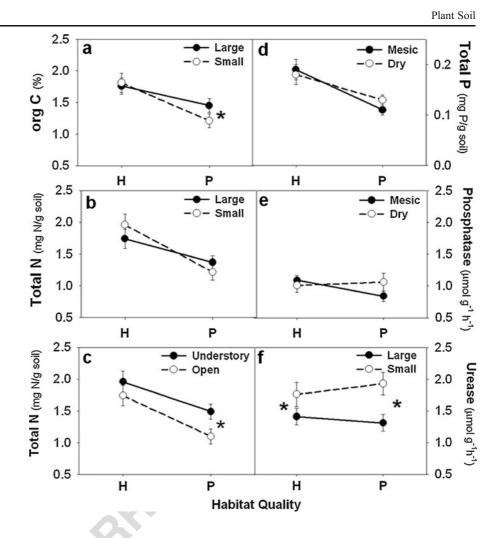
384 Allen and Schlesinger 2004). Moreover, greater levels of plant production (e.g. litter production) also stimu-385late microbial enzymatic activity (Zak et al. 2003; 386 Allen and Schlesinger 2004). B-glucosidase, urease 387 and acid phosphatase are hydrolases involved in the 388 decomposition of complex compounds. In particular, 389 390 β-glucosidase has a key role in the C cycle, it is responsible for the transformation of large chains of 391carbohydrates into assimilable sugars (Eivazi and 392393 Zakaria 1993). Thus a decrease in ß-glucosidase activity has negative effects on the activity of other 394enzymes (Sardans and Penuelas 2005). These findings 395 396 together with the patchy distribution of plants in Mediterranean gypsum soils support the idea that plant 397 cover clumps in dry ecosystems function as resource 398 399 islands with milder living conditions arranged in a barren matrix of bare soil (Maestre and Cortina 4002002; Goberna et al. 2007). 401

between high and poor habitat quality (bar color). Abbreviations are: H, high-habitat quality; P, poor-habitat quality; L, large fragment; S small fragment; M, mesic treatment (watered plants); D, dry treatment (non-watered treatments). See text for details

The lack of direct effect of water availability could 402 be explained by the different time scale at which this 403driver can impact on ecosystem properties. For exam-404 ple, in our study case, the 2 years of manipulative 405 changes in water availability contrasts with the long 406 term processes associated with loss of habitat quality. 407 Nutrient availability usually shows a lagged response 408 to climatic variations, sometimes taking even decades 409to respond to environmental variation in the case of 410 so-called slow variables (Reynolds et al. 2007). How-411 ever, enzymatic activities are rapid soil functional 412 surrogates and therefore short-term effects of our wa-413ter treatment can be expected. We did not detect these 414 effects for the different water treatments due to our 415 experiment mimicking either a mild or too short 416drought or a drought not affecting soils at the most 417responsive time of the year; it must be noted, however, 418that our drought simulation was guided both in extent 419

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Fig. 4 Significant interactions between Habitat Quality and global change drivers (Fragmentation, Water availability and Microhabitat). Values are mean \pm SE in each treatment. Graphs only show significant interactions between factors. An *asterisk* indicates significant differences (at *p*<0.05) between levels of a factor. Abbreviations are: H, high-habitat quality; P, poor-habitat quality



and timing by climate change scenarios and not by the 420 responsiveness of soil biota. This explanation is sup-421ported by results from other studies showing that 422drought significantly decreased soil enzymatic activity 423424 when more intense rainfall reductions were simulated (Sardans and Penuelas 2005) or when long-term rain-425fall variations were explored (Li and Sarah 2003). 426427 Likewise, fragmentation did not have a significant direct effect on any of the response variables. Accord-428 429 ing to the literature, microbial communities are, in general, not sensitive to habitat fragmentation and 430habitat size (Rantalainen et al. 2005 and 2008). How-431 ever, this does not mean that fragmentation is irrele-432vant for soil functioning. We found that fragmentation 433 indirectly affected soil performance (e.g. the effects of 434loss of habitat quality on nutrient availability were 435exacerbated in small fragments). Therefore, studying 436the effect of habitat fragmentation on soil features and 437

performance is critical, especially in combination with 438 other global change drivers. 439

Interactive effects of global change drivers 440

As hypothesised, habitat quality, fragmentation and 441water availability interactively affected nutrient 442 availability and microbial activity of Mediterranean 443 gypsum soils (Sala et al. 2000; Brook et al. 2008; 444 Matesanz et al. 2009; Pias et al. 2010). First of 445 all, we found that the negative impact of habitat 446 quality loss on total organic C and total N was 447exacerbated in small fragments, which is relevant 448 to predict the final outcome of land degradation on 449 ecosystem functioning since, both drivers usually act 450together (Schleuning et al. 2008). Second, we found that 451the reduction of total N from high- to poor- habitat 452quality sites was greater in open areas than under the 453

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understory of C. hyssopifolia. This result agrees with 454other studies showing that microhabitat heterogeneity 455456modulates the impact of global change drivers such as loss of habitat quality (Maestre and Reynolds 2006). 457Furthermore, given that soil nutrient heterogeneity 458exerts a strong influence on the development of plant 459individuals and communities (Hodge et al. 2000; Day et 460 al. 2003), we can expect ecological processes mediated 461 462by environmental heterogeneity (such as plant distribution or plant-plant interactions) to be indirectly affected 463by habitat quality loss. Finally, we found an interaction 464 between habitat quality and water availability. Contrary 465 to our expectations, we did not find significant differ-466 ences in total P and phosphatase activity between water-467 ing treatments. Our results contrast with other studies 468 showing that enzymatic activity is correlated with soil 469water availability in semiarid (Kramer and Green 2000) 470 and dry Mediterranean soils (Li and Sarah 2003; Sardans 471472and Penuelas 2004, 2005).

Fragmentation affects plant survival due to de-473474creased genetic variation and increased inbreeding 475(Ellstrand and Elam 1993; Fischer et al. 2003). This has been also suggested by Matesanz et al. (2009) in a 476477 previous study in the same system, where the interac-478tion between habitat quality and fragmentation affected survival and relative growth of C. hyssopifolia. 479 This reduction in plant survival, and therefore in plant 480 481 cover, reduces organic matter content in the soil and could, in turn, affects soil microbial activity, in small 482 fragments. According to these results, fragmentation 483did not have a significant direct effect on soils features 484and performance, but it modulated the effect of habitat 485quality through synergistic interactions having an in-486direct effect on soil properties mediated by plant cover 487 decline. 488

489 Conclusions

Our results highlight the importance of considering 490491 several drivers simultaneously to forecast realistic ecosystem responses to global change impacts (Sala et al. 4922000; Matesanz et al. 2009). Each driver operates on 493different time scales: year to year change for water 494availability versus decades for habitat quality loss and 495fragmentation. This different time scale of the drivers 496497 could explain the greater effect of habitat quality on soils properties, which could be exacerbated by the 498interactive effect of habitat fragmentation over a long 499

time scale. Moreover, there are feedbacks between 500 plant and microbial activity so cumulative effects of 501 drivers affecting plant productivity and microbial activity and interactions among them can be expected in 503 the long-term and could accelerate the degradation of 504 Mediterranean gypsum habitats. 505

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- Q1. Please check capturing of Affiliations 1–3 if appropriate.
- Q2. The citation "Garcia et al. 2000" (original) has been changed to "Garcia et al. 2002". Please check if appropriate.

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